Lysate Assay:
The first step was to make a cytophaga liquid media suitable for the growth of *Flavobacterium psychrophilum*. The recipe for the liquid media is identical to the recipe used to make cytophaga agar plates, minus the agar. All lysate assay tests were run with 20 mL cytophaga media samples. The samples were inoculated from a -80°C frozen glycerol stock of *F. psychrophilum* and then incubated at 15°C for 48-72 hours.

An overnight culture of EcnB-BL21 constructs were used to inoculate a preinduction culture consisting of LB and chloramphenicol (25µg/mL). The cultures were incubated at 37°C for 6 hours and induced with L-arabinose (1% w/w). After 6 more hours of incubation at 37°C, 125 mL of culture were centrifuged at 5000g for 10 minutes. The pellets were resuspended in 1 mL phosphate buffer saline solution and homogenized (4) for 10 minutes. The lysates were centrifuged at 10000g for 10 minutes. Samples were placed either on ice or at 4°C throughout the lysis process. The supernatants were added to the *Flavobacterium* cultures. Optical density readings were performed using a spectrophotometer set at OD600. Readings were performed over 12 hours.